

Physical Characterization and Droplet Dispersion of Sneeze or Cough Ejecta: A Review

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Abstract - Physical properties of a sneeze or cough ejecta reported in the open literature are reviewed to characterize the two-phase flow (exhaled air and mucosalivary droplets). Sneeze or cough ejecta are studied as a two-phase buoyant puff at droplet sizes of 0.15-100 μ m. Then droplet dispersion by a sneeze or cough ejecta to quasi-static environments is studied, from a physics view, to exploring the possibility of airborne transmission of viruses as SARS-CoV-2. Nozzle stokes numbers are analyzed at three Reynolds numbers, which correspond to the representative velocities of exhaled air (10, 15, and 20 m/s). Calculations show particles smaller than 5 μ m are the critical case for airborne transmission, whereas particles less than 19 μ m are affected by turbulent motions and remain airborne for long periods.

Key Words: sneeze or cough ejecta, two-phase puff, droplet dispersion, nozzle Stokes number, airborne.

1. INTRODUCTION

In the open literature, there is much information on the physical characteristics of human respiratory droplets, which are focused on talking, coughing, and sneezing. The experimental data coincide with many droplets generated in the sneeze ejecta, and there are similar droplets distributions on talking, coughing, and sneezing. Some authors, as Zayas et al. [1], found that there is no significant correlation between droplet size and cough frequency, age, and gender. However, there are controversies in the droplet sizes, exhaled air velocity, and droplet velocity by the measurement methods. The physical characteristics of air and droplets exhaled by sneeze, cough, or talking are relevant in the droplet dispersion. Droplet dispersion on controlled conditions allows understanding airborne transmission of viruses as SARS-CoV-2, which has caused 1.8 million deaths around the world [2] until January 2021. Therefore, the main objective of the review is to study physical characteristics of sneeze or cough ejecta and droplet dispersion in indoor environments. The review is focused on the physical mechanisms of droplet dispersion of two-phase buoyant puff. The sneeze or cough ejecta and the nozzle Stokes number in droplets of 0.15-50 µm are studied to understand the physical mechanisms of a potential airborne transmission of viruses as SARS-CoV-2 from a physics view.

2. PHYSICAL CHARACTERISTICS

There are many methods on droplet size and velocities of droplet or exhaled air by zneeze, cough or breathing. According to Zhang et al. [3], the Impaction Methods (IM) may cause particles to spread, splash, or finger and distort the correct particle size if identified by microscopy. Then other methods used are the Aerodynamic Particle Sizer (APS), Cough Aerosol Sampling System (CASS), High-Speed (HS) camera/video, Hot-Wire Anemometry (HWA) Interferometric Mie Imaging (IMI), Laser (Particle Size Analyser (PSA), Light Scattering (LS), and Diffraction System (DS)), Optical Particle Counter (OPC), Optical Particle Spectrometer (OPS), Particle Image Velocimetry (PIV), Scanning Mobility Particle Sizer (SMPS), Schlieren PIV (S-PIV), Shadowgraph Imaging (S-Imag), and Spirometer. The measurement methods, droplet size (D_p) , and air/droplet velocity (*u*) on talking, coughing, and sneezing, are summarized in Table 1. Most experimental data show droplet sizes between 0.15-100 µm, average velocities of exhaled air of 4.5-21.25 m/s, and average velocities of droplets between 5-22 m/s.

Other characteristics of the sneeze or cough ejecta are the mouth opening area, which is based on human cough captured on photographs, and geometric plane shapes are approached as a semi-circular section (Gupta et al. [4]; Busco et al. [5]) or a rectangular sheet-like (Dbouk & Drikakis [6]). The semi-circular section approximation is considered as a characteristic length (equivalent diameter of an exit nozzle). This equivalent diameter is calculated from Gupta et al. [4] data (mouth opening area of $3.37-4 \text{ cm}^2$), which agrees to 3.4 cm^2 reported by Bourouiba et al. [7]. Calculation of the equivalent diameter is 0.021-0.023 m, which agrees with the mouth diameter (D_{em}) of 0.02 m used in many numerical simulations.

Table -1: Droplet size and velocity of airflow/droplet.

Author	Method	Dp (µm)	u (m/s)	
[8]	APS	0.62-15.9 and 0.58-5.42 ^{dn}	na	
[9]	APS	0.5-20	na	
[10]	CASS	< 3 (70%)	na	
[11]	HS camera	7-100	na	



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[12]	HS video	na	> 6 ^{droplet}		
[13]	HS camera	160-1000	14droplet, maxv		
[14]	HS camera	na	12-15 ^{droplet, maxv} , 5 ^{droplet, ps}		
[15]	IM (solid)	1-200 ^{av}	na		
[16]	IM (liquid)	50-860	na		
[17]	IM (solid)	< 1	na		
[18]	IM (solid)	55 ^{av}	na		
[19]	IM (solid)	< 1	na		
[20]	IM (solid)	$50-100^{av}$ and 5^{minv}	na		
[21]	Laser PSA	360 ^{av, ud} and 74.4 ^{av, bd}	na		
[22]	Laser LS	20-500	na		
[23]	OPC	0.15-0.5 and 0.15-0.199 ^{ps}	na		
[24]	OPC	0.3-5	na		
[25]	OPC	0.3499 (82%)	na		
[26]	PIV	na 6-22droplet, 11 2droplet, av			
[27]	PIV	na	10.6 ^{air, female} , 15.3 ^{air, male}		
[28]	PIV-IMI	13.5 ^{av}	11.7 ^{air}		
[29]	PIV	na	1.15-28.8 ^{air} , 10.2 ^{av}		
[30]	PIV-HWA	na	3.05 ^{air, jet} centre (1 17 ^{av, peak})		
[31]	SMPS-OPS	< 5 (99%)	15.03-19.55 ^{air, **}		
[32]	S-PIV	na	8air, maximum av		
[33]	S-Imag	na	4.5 ^{air, maxv}		
[4]	Spiromete r	na	4.75-21.25 ^{air,*}		

* value calculated considering the information provided in the paper.

 ** value calculated considering a mouth opening area of 3.37-4 \mbox{cm}^2

av average value

bd bimodal distribution

dn droplet nuclei

minv minimum value

maxv maximum value

ps predominant size

ud unimodal distribution

The time of sneeze or cough expulsion phase lasted approximately 200-300 ms (Bahl et al. [14]; Bourouiba et al. [7]; Scharfman et al. [13]), although Busco et al. [5] and Zayas et al. [1] have reported average times of 500 ms and 700 ms, respectively. The average cone spreading angles are $24 \pm 3^{\circ}$ (Dudalski [30]), 25° (Gupta et al. [4], calculated by subtracting the directional angles), and $23.9^{\circ} \pm 3.48^{\circ}$ (Tang et al. [32]). These angles agree to the universal spreading angle of 23.8° of the jet dispersion theory (Cushman-Roisin [34]). The potential core (Fig. 1) has not been presented in experiments, although a slight representation is shown in the velocity contour of a cough onset by Tang et al. [32]. Exhaled air temperatures on a sneeze or cough ejecta of 30-33°C are reported by Tang et al. [33]. These values agree to exhaled breath temperatures (33.8°C, 33.2°C±1.3°C, and 31.4-35.4°C) by Roberge et al. [35], Bijnens et al. [36], and Mansour et al. [37], respectively. Experimental measurements of exhaled breath RH showed variations of 65.0-88.6% and 41.9-91.0%, according to Mansour et al. [37]. However, Niesters et al. [38] and the ScienceBits website [39] reported values close to saturation and 95%, respectively.

3. PHYSICS OF DROPLETS DISPERSION

Fluids expelled on a sneeze or cough ejecta are mainly composed of exhaled air and mucosalivary droplets of various sizes. Sneezing or coughing is considered as a twophase buoyant puff, which could study in two-time stages: stage 1) during the expulsion of fluids (air and droplets) and stage 2) post-expulsion of fluids. In stage-1 (fluids expelled), the fluid momentum is the dominant mechanism on the jet dispersion, and the physical phenomenon is approached as a one-phase turbulent jet dispersion. Froude number $(Fr=u_i/[(\rho_i-\rho_a)/\rho_a)gD_{em}])$ and Richardson number $(Ri=\rho_i-\rho_a)$ $\rho_a/gD_{em}/\rho_i u_i^2$) are calculated for the two-phase jet, as shown in Table 2. The subscripts *j* and *a* are the jet (exhaled air) and the atmospheric air, respectively. Computations consider properties of exhaled air at 90% RH, which are calculated with data provided by Tsilingiris [40]. Results show Fr >> 1and Ri << 0.1, which match criteria to determine if a jet dispersion is dominated by the initial momentum (Bricard & Friedel [41]). Later a turbulent cloud is formed (stage 2), and the jet trajectory of the centerline is curved by density difference as reported by Bourouiba et al. [7] and Wei & Li [42], and it's shown in nature video [43]. The buoyant dominates the physical phenomenon by velocity decay and warmer temperature. Then turbulent cloud moves along the cone in a meandering motion (see Fig. 1), which is approximated as a puff in a stratified environment. The turbulent cloud loses buoyancy and will move until the bulk cloud velocity is zero. A wide range of eddy sizes into de cloud disperse droplets and contribute to the cloud expansion, as seen in the nature video [43]. The average turbulence intensity reported by Dudalski [30] of a sneeze or cough is 8.9±3.9% at 45 times the equivalent diameter ($D_{em} = 0.0217$ m).

The physiochemical phenomena of the ejection of mucosalivary fluid are complex by the multiphase nature of the flow and the inhomogeneous liquid phase formed by droplets of different sizes, ligaments, bags of mucosaliva, and pearls (Scharfman et al. [13]). The human saliva presents a marked non-Newtonian flow effect (Haward et al. [44]), and there are salt/electrolytes (Johnson & Morawska [9]; Liu et al. [45]). In the sneeze or cough ejecta, droplets larger than 100 µm follow a projectile motion. Particles fall out by gravity as a rainout phenomenon with no appreciable vaporization (Wells [46]; Wei & Li [42]; Li et al. [47]; Yan et al. [48]), while droplet sizes less than 100 μ m remain airborne (Duguid [15]; Li et al. [47]). Discussion of the phenomenon is conducted to droplets smaller than 100 µm because they are affected by flow dynamics of buoyant puff, and the evaporation rate is relevant. On small droplets ($D_p <$ $100 \,\mu\text{m}$), the evaporation is complex by the salt/electrolytes content in the saliva, the relative humidity of the exhaled air, and the droplet size. The salt/electrolytes content affects the droplet evaporation (Johnson & Morawska [9]; Zhang [49]), and the drying time increases around 20% (Liu et al. [45]), despite the physical properties of saliva are close to the water with 99.5% water, 0.3% proteins, and 0.2% inorganic/trace substances (Schipper et al. [50]). The relative humidity of the exhaled air has a strong effect on the drying time and increases almost 7-fold as humidity increase up to 90% (Li et al. [47]; Bhardwaj & Agrawal [51]). Then energy and mass transport are poor because liquid-phase and gas-phase could be considered in quasi-equilibrium. According to Wei & Li [42], RH is a key parameter to the droplet evaporation and spread of the virus. Finally, droplet evaporation increases as droplet size decrease by the enlarged specific surface area for heat and mass transfer. However, the evaporation rate of the microdroplet ($D_p \le 2$ μm) is slowed down because the droplet size is comparable to the mean free path of air molecules, according to Hołyst et al. [52].

A physics view of a sneeze or cough ejecta on the environment could help to understand the possibility of airborne transmission of viruses. Then a parameter that provides information on particle dispersion by its inertial response time is the Stokes number (*Stk*). According to Lau & Nathan [53], this parameter has a strong impact on the particle concentration and the subsequent evolution of the two-phase jet. There are four types of Stokes numbers (nozzle Stk, turbulent Stk, Kolmogorov Stk, and acceleration Stk), as described by Kennedy & Moody [54]. The nozzle Stokes number (*Stk_n*) has an overall representation of the phenomenon because it is based on the Stokes law and the convective time scale of the mean flow. So, the nozzle Stokes number (*Stk_n* = $C_c \rho_p D_p 2u_i / 18 \mu D_{em}$) is calculated with the equivalent diameter of the mouth opening (D_{em}) as a nozzle diameter. The subscript p is the particle (droplet), and C_c is the Cunningham coefficient caused by slippage. Droplet diameters are normalized (D_p/D_o) to a droplet size (D_o) of 5

μm, which remain suspended in the air, according to WHO [55]. Computations are conducted at droplet diameters up to 50 μm, where particles remain airborne. Three Reynolds numbers of exhaled air (Re_{ea}) are considered, according to experimental velocities (10, 15, and 20 m/s) presented in §2, at an average temperature of 33°C (see §2). The thermodynamic conditions considered for mucosalivary ejecta are presented in the notes of Table 2, and results are shown in Fig. 2.



Fig -1: Physical representation of a sneeze or cough ejecta on quasi-static environments.

Table -2: Dimensionless parameters of the two-phase jetat two exhaled air temperatures.

Re _{ea}	<i>T_{ea}</i> (°C)		Fr		Ri*1E5	
13877	30	35	130.2	104.3	6.1	9.6
20815	30	35	195.3	156.5	2.7	4.3
27754	30	35	260.4	208.6	1.5	2.4

environmental air at 25°C and 101 kPa

 $D_{em} = 0.022 \text{ m}$

$$g = 9.8 \text{ m/s2}$$

exhaled air properties at 90% of RH and 101 kPa



Fig -2: Nozzle Stokes number of a sneeze or cough ejecta.

Fig. 2 shows droplets smaller than 5 μ m ($D_p/D_p = 1$) are in a mechanical quasi-equilibrium with the flow and are transported by the airflow as a gas-phase ($Stk_n \ll 1$). This result agrees with experiments conducted by van Doremalen et al. [56], who find SARS-CoV-2 remains viable in particles smaller than 5 µm for approximately 3 hrs with a half-life of 1.1-1.2 h. In the case of $Stk_n < 1$, the mechanical quasiequilibrium is valid because particle dispersion is high, and slip velocity between particles and gas phase is low (Sun et al. [57]). Then droplets smaller than 19 μ m ($D_p/D_o < 3.8$) are transported by airflow, and they remain airborne for long periods. Particles smaller than 50 μ m ($D_p/D_o = 10$) respond to turbulent motions and are carried by the stream (Stk_n between 1-9). The influence of airflow on droplets between 50-100 μ m is weak (10 < *Stk_n* < 100), although particles are still carried by airflow. Droplets larger than $100 \,\mu m (D_p/D_o \ge 10^{-3})$ 20) are not affected by airflow because the nozzle Stokes number is higher than 100 ($Stk_n >> 1$). These results show particles smaller than 19 µm remain airborne in the environment for long periods, and the limit value is $50 \mu m$. The particle dispersion is focused on a quasi-static environment under controlled conditions as confined spaces. Therefore, the airborne transmission of viruses in a quasistatic environment is possible from a physics view. The risk of infections in non-ventilated rooms is high, even if the distance is too large for direct transmission of the virus (Riediker & Tsai [58]).

4. CONCLUSION

Fluids expelled on a sneeze or cough could be studied in twotime stages, as a two-phase buoyant puff. In stage-1 (during the expulsion of fluids), the fluid momentum is the dominant mechanism on the jet dispersion. In stage-2 (post-expulsion of fluids), the buoyancy effect dominates the physical phenomenon. Nozzle Stokes numbers show particle sizes less than 5 µm remain airborne as a gas-phase ($Stk_n << 1$), which is the critical case to airborne transmission of viruses. Particles smaller than 19 µm ($Stk_n < 1$) are transported by airflow and remain airborne in the environment for long periods. Droplets between 50-100 µm are still carried by airflow, but airflow influence is weak ($10 < Stk_n < 100$), whereas droplets larger than 100 µm ($D_p/D_o \ge 20$) are not affected by airflow.

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REFERENCES

 Zayas, G., Chiang, M.C., Wong, E., MacDonald, F., Lange, C.F., Senthilselvan, A., King, M., 2012. Cough aerosol in healthy participants: fundamental knowledge to optimize droplet-spread infectious respiratory disease management. Pulm. Med. 12, 1-11. http://www.biomedcentral.com/1471-2466/12/11.

- [2] World Health Organization (WHO), 2020. Weekly epidemiological update - 5 January 2021. https://www.who.int/publications/m/item/weeklyepidemiological-update---5-january-2021.
- [3] Zhang, H., Lia, D., Xie, L., Xiao, Y., 2015. Documentary Research of Human Respiratory Droplet Characteristics. Procedia Engineering 121, 365-1374. https://doi.org/10.1016/j.proeng.2015.09.023.
- [4] Gupta, J.K., Lin, C.-H., Chen, Q., 2009. Flow dynamics and characterization of a cough. Indoor Air 19, 517-525. https://doi.org/10.1111/j.1600-0668.2009.00619.x.
- [5] Busco, G., Yang, S.R., Seo, J., Hassan, Y.A., 2020. Sneezing and asymptomatic virus transmission. Phys. Fluids 32, 073309. https://doi.org/10.1063/5.0019090.
- [6] Dbouk, T., Drikakis, D., 2020. On coughing and airborne droplet transmission to humans. Phys. Fluids 32, 053310. https://doi.org/10.1063/5.0011960.
- Bourouiba, L., Dehandschoewercker, E., Bush J.W.M.,2014. Violent expiratory events: on coughing and sneezing. J. Fluid Mech. 745, 537-563. http://dx.doi.org/10.1017/jfm.2014.88.
- [8] Yang, S., Lee, G.W.M., Chen, C-M., Wu, C-C., Yu, K-P., 2007. The Size and Concentration of Droplets Generated by Coughing in Human Subjects. J. Aerosol Med. 20, 484-494. https://doi.org/10.1089/jam.2007.0610.
- [9] Johnson, G. R., Morawska, L., 2009. The mechanism of breath aerosol formation. J. Aerosol. Med. Pulm. Drug Deliv. 22, 229-237. https://doi.org/10.1089/JAMP.2008.0720.
- [10] Wainwright, C.E., France, M.W., O'Rourke, P., Anuj, S., Kidd, T.J., Nissen, M.D., Sloots, T.P., Coulter, C., Ristovski, Z., Hargreaves, M., Rose, E.R., Harbour, C., Bell, S.C., Fennelly, K.P., 2009. Cough-generated aerosols of Pseudomonas aeruginosa and other Gram-negative pathogen from patients with cystic fibrosis, Thorax 64, 926-931. http://dx.doi.org/10.1136/thx.2008.112466.
- [11] Jennison M.W., 1942. Atomizing of mouth and nose secretions into the air as revealed by high-speed photography. Aerobiology 17, 106-128.
- [12] Nishimura, H., Sakata, S., Kaga, A., 2013. A New Methodology for Studying Dynamics of Aerosol Particles in Sneeze and Cough Using a Digital High-Vision, High-Speed Video System and Vector Analyses. PLoS ONE 8, e80244. https://doi.org/10.1371/journal.pone.0080244.
- [13] Scharfman, B.E., Techet, A.H., Bush, J.W.M., Bourouiba, L., 2016. Visualization of sneeze ejecta: steps of fluid fragmentation leading to respiratory droplets. Exp.

Fluids 57, 1-9. https//doi.org/10.1007/s00348-015-2078-4.

- [14] Bahl, P., de Silva, C.M., Chughtai, A.A., MacIntyre, C.R., Doolan, C., 2020. An experimental framework to capture the flow dynamics of droplets expelled by a sneeze. Exp. Fluids 61, 1-9. https://doi.org/10.1007/s00348-020-03008-3.
- [15] Duguid, J.P., 1946. The size and the duration of aircarriage of respiratory droplets and droplet-nuclei. J. Hyg. 44, 471-479. doi:10.1017/S0022172400019288.
- Buckland, F.E., Tyrrell, D.A.J., 1964. Experiments on spread of colds: 1. Laboratory studies on dispersal of nasal secretion. J. Hyg. 62, 365-377. https://doi.org/10.1017/S0022172400040080.
- [17] Gerone, P.J., Couch, R.B., Keefer, G.V., Douglas, R.G., Derrenba, E.B., Knight, V., 1966. Assessment of experimental and natural viral aerosols. Bacteriol. Rev. 30, 576-588.
- [18] Loudon, R.G., and Roberts, R.M., 1967. Droplet expulsion from respiratory tract. Am. Rev. Respir. Dis. 95, 435-442. https://doi.org/10.1164/arrd.1967.95.3.435.
- [19] Papineni, R.S, Rosenthal, F.S., 1997. The size distribution of droplets in the exhaled breath of healthy human subjects. J. Aerosol Med. 10, 105-116. doi:10.1089/jam.1997.10.105.
- [20] Xie, X., Li, Y., Sun, H., Liu, L., 2009. Exhaled droplets due to talking and coughing. J. R. Soc. Interface 6, S703–S714. doi:10.1098/rsif.2009.0388.focus.
- [21] Han, Z.Y., Weng, W.G., Huang, Q.Y., 2013. Characterizations of particle size distribution of the droplets exhaled by sneeze. J. R. Soc. Interface 10, 1-11. http://dx.doi.org/10.1098/rsif.2013.0560.
- [22] Anfinrud, P., Stadnytskyi, V., Bax, C.E., Bax, A., 2020. Visualizing Speech-Generated Oral Fluid Droplets with Laser Light Scattering. N. Engl. J. Med. 382, 2061-2063. https://doi.org/10.1056/NEJMc2007800.
- [23] Edwards, D.A., Man, J.C., Brand, P., Katstra, J.P., Sommerer, K., Stone, H.A., Nardell, E., Scheuch, G., 2004. Inhaling to mitigate exhaled bioaerosols. Proc. Natl. Acad. Sci. USA 101, 17383-17388. doi:10.1073/pnas.0408159101.
- [24] Fabian, P., McDevitt, J.J., DeHaan, W.H., Fung, R.O.P., Cowling, B.J., Chan, K.H., Leung, G.M., Milton, D.K., 2008. Influenza virus in human exhaled breath: an observational study. PLoS ONE 3, e2691. doi:10.1371/journal.pone.0002691.
- [25] Fabian, P., Brain, J., Houseman. E.A., Gern, J., Milton, D.K., 2011. Origin of exhaled breath particles from healthy

and human rhinovirus-infected subjects. J. Aerosol Med. Pulm. Drug Deliv. 24, 137-147. doi:10.1089/jamp.2010.0815.

- [26] Zhu, S., Kato, S., Yang, J.H., 2006. Study on transport characteristics of saliva droplets produced by coughing in a calm indoor environment. Build. Environ. 41, 691-1702. https://doi.org/10.1016/j.buildenv.2005.06.024.
- [27] Kwon, S.-B., Park, J., Jang, J., Cho, Y., Park, D.-S., Kim, C., Bae, G.-N., Jang, A., 2012. Study on the initial velocity distribution of exhaled air from coughing and speaking. Chemosphere 87, 1260-1264. doi: 10.1016/j.chemosphere.2012.01.032.
- [28] Chao, C.Y.H., Wan, M.P., Morawska, L., Johnson, G.R., Ristovski, Z.D., Hargreaves, M., Mengersen, K., Corbett, S., Li, Y., Xie, X., Katoshevski. D., 2009. Characterization of expiration air jets and droplet size distributions immediately at the mouth opening. J. Aerosol Sci. 40, 122-133. https://doi.org/10.1016/j.jaerosci.2008.10.003.
- [29] VanSciver, M., Miller, S., Hertzberg, J., 2011. Particle Image Velocimetry of Human Cough. Aerosol Sci. Tech. 45, 415-422. https://doi.org/10.1080/02786826.2010.542785.
- [30] Dudalski, N., 2018. Experimental measurements of human cough airflows from healthy subjects and those infected with respiratory viruses. A Master thesis submitted in Master of Engineering Science, University of Western Ontario. https://ir.lib.uwo.ca/etd/5965.
- [31] Lee, J., Yoo, D., Ryu, S., Ham, S., Lee, K., Yeo, M., Min, K., Yoon, C., 2019. Quantity, size distribution, and characteristics of cough-generated aerosol produced by patients with an upper respiratory tract infection. Aerosol Air Qual. Res. 19, 840-853. https://doi.org/10.4209/aaqr.2018.01.0031.
- [32] Tang, J.W., Liebner, T.J., Craven, B.A., Settles, G.S., 2009. A schlieren optical study of the human cough with and without wearing masks for aerosol infection control. J. R. Soc. Interface 6, S727-S736. https://doi.org/10.1098/rsif.2009.0295.focus.
- [33] Tang, J.W., Nicolle, A.D., Klettner, C.A., Pantelic, J., Wang, L., Suhaimi, A.B., Tan, A.Y., Ong, G.W., Su, R., Sekhar, C., Cheong, D.D., Tham, K.W., 2013. Airflow Dynamics of Human Jets: Sneezing and Breathing - Potential Sources of Infectious Aerosols. PLoS ONE 8, e59970. https://doi.org/10.1371/journal.pone.0059970.
- [34] Cushman-Roisin. B., 2019. Environmental Fluid Mechanics. In: John Wiley & Sons (to be published), pp. 155.https://www.dartmouth.edu/~cushman/books/EF M-old.html.

- [35] Roberge, R.J., Bayer, E., Powell, J.B., Coca, A., Roberge, M.R., Benson, S.M., 2010. Effect of Exhaled Moisture on Breathing Resistance of N95 Filtering Facepiece Respirators. Ann. Occup. Hyg. 54, 671-677. https://doi.org/10.1093/annhyg/meq042.
- [36] Bijnens, E., Pieters, N., Dewitte, H., Cox, B., Janssen, B. G., Saenen, N., Dons, E., Zeegers, M.P., Int Panis, L., Nawrot, T. S., 2013. Host and environmental predictors of exhaled breath temperature in the elderly. BMC public health 13, 1226. https://doi.org/10.1186/1471-2458-13-1226.
- [37] Mansour, E., Vishinkin, R., Rihet, S., Saliba, W., Fish, F., Sarfati, P., Haick, H., 2020. Measurement of temperature and relative humidity in exhaled breath. Sensor. Actuat. B-Chem. 304, 127371. https://doi.org/10.1016/j.snb.2019.127371.
- [38] Niesters, M., Mahajan, R., Olofsen, E., Boom, M., Garcia del Valle, S., Aarts, L., Dahan, A., 2012. Validation of a novel respiratory rate monitor based on exhaled humidity. Brit. J. of Anaesth. 109, 981-989. https://doi.org/10.1093/bja/aes275.
- [39] ScienceBits website, 2020. Condensation of your exhaled breath. http://www.sciencebits.com/exhalecondense.
- [40] Tsilingiris, P.T., 2008. Thermophysical and transport properties of humid air at temperature range between 0 and 100 °C. Energ. Convers. Manage. 49, 1098-1110. https://doi.org/10.1016/j.enconman.2007.09.015.
- [41]
 Bricard, P., Friedel, L., 1998. Two-phase jet dispersion. J.

 Hazard.
 Mater.
 59,
 287-310.

 https://doi.org/10.1016/S0304-3894(97)00159-3.
- [42] Wei, J., Li, Y., 2015. Enhanced spread of expiratory droplets by turbulence in a cough jet. Build. Environ. 93, 86-96. http://dx.doi.org/10.1016/j.buildenv.2015.06.018.
- [43] Nature video, 2016. The physics of sneeze. https://www.youtube.com/results?search_query=the+p hysics+of+sneeze.
- [44] Haward, S.J., Odell, J.A., Berry, M., Hall, T., 2011. Extensional rheology of human saliva. Rheol. Acta 50, 869-879. https://doi.org/10.1007/s00397-010-0494-1.
- [45] Liu, L., Wei, J., Li, Y., Ooi, A., 2017. Evaporation and dispersion of respiratory droplets from coughing. Indoor Air 27, 179-190. https://doi.org/10.1111/ina.12297.
- [46] Wells, W.F., 1934. On air-borne infections: study II. Droplets and droplet nuclei. Am. J. Epidemiol. 20, 611– 618.

https://doi.org/10.1093/oxfordjournals.aje.a118097.

- [47] Li, X., Shang, Y., Yan, Y., Yang, L., Tu, J., 2018. Modelling of evaporation of cough droplets in inhomogeneous humidity fields using the multi-component Eulerian-Lagrangian approach. Build. Environ. 128, 68-76. https://doi.org/10.1016/j.buildenv.2017.11.025.
- [48] Yan, Y., Li, X., Tu, J., 2019. Thermal effect of human body on cough droplets evaporation and dispersion in an enclosed space. Build. Environ. 148, 96-106. https://doi.org/10.1016/j.buildenv.2018.10.039.
- [49] Zhang, T., 2011. Study on Surface Tension and Evaporation Rate of Human Saliva, Saline, and Water Droplets. Master thesis of Science in Mechanical Engineering. West Virginia University. https://researchrepository.wvu.edu/etd/2271/.
- [50] Schipper, R.G., Silletti, E., Vingerhoeds, M.H., 2007. Saliva as research material: biochemical, physicochemical and practical aspects. Arch. Oral Biol. 52, 1114-1135. https://doi.org/10.1016/j.archoralbio.2007.06.009.
- [51] Bhardwaj, R., Amit Agrawal, A., 2020. Likelihood of survival of coronavirus in a respiratory droplet deposited on a solid surface. Phys. Fluids 32, 061704. https://doi.org/10.1063/5.0012009.
- [52] Hołyst, R., Litniewskia, M., Jakubczyk, D., 2017. Evaporation of liquid droplets of nano- and micro-meter size as a function of molecular mass and intermolecular interactions: experiments and molecular dynamics simulations. Soft. Matter 13, 5858-5864. https://doi.org/10.1039/C7SM00804J.
- [53] Lau, T.C.W., Nathan, G.J., 2014. Influence of Stokes number on the velocity and concentration distributions in particle-laden jets. J. Fluid Mech. 757, 432-457. https://doi.org/10.1017/jfm.2014.496.
- [54] Kennedy, I.A., Moody, M.H., 1998. Particle dispersion in a turbulent round jet. Exp. Therm. Fluid Sci. 18, 11-26. https://doi.org/10.1016/S0894-1777(98)10009-2.
- [55] World Health Organization (WHO), 2020. Modes of transmission of virus causing COVID-19: implications for IPC precaution recommendations. Scientific brief, first published on 29 March 2020, updated on 9 July based on updated scientific evidence. https://www.who.int/newsroom/commentaries/detail/modes-of-transmission-ofvirus-causing-covid-19-implications-for-ipc-precautionrecommendations.
- [56] van Doremalen, N., Bushmaker, T., Morris, D. H., Holbrook, M. G., Gamble, A., Williamson, B.N., Tamin, A., Harcourt, J. L., Thornburg, N. J., Gerber, S. I., Lloyd-Smith, J. O., de Wit, E., Munster, V. J., 2020. Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1. N. Engl. J. Med., 382, 1564-1567. DOI: 10.1056/NEJMc2004973.



- [57] Sun, G., Hewson, J.C., Lignell, D.O., 2017. Evaluation of Stochastic Particle Dispersion Modeling in Turbulent Round Jets. Int. J. Multiphas. Flow 89, 108-122. https://doi.org/10.1016/j.ijmultiphaseflow.2016.10.00 5.
- [58] Riediker, M., Tsai, D-H., 2020. Estimation of viral aerosol emissions from simulated individuals with asymptomatic to moderate coronavirus disease 2019.
 JAMA Network Open 3, e2013807. https://doi.org/10.1001/jamanetworkopen.2020.1380
 7.